

BactoReal[®] Kit *Chlamydiaceae*

Manual

For use with the

- ABI PRISM[®] 7500 (Fast)
- Mx3005P[®]
- LightCycler[®] 480



For research only, not for diagnostic use



DVEB03111, DVEB03113



100



DVEB03151, DVEB03153



50



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Index

1. Product description	3
2. Pathogen information	3
3. Principle of real-time PCR.....	3
4. General Precautions	3
5. Contents of the Kit.....	4
5.1. BactoReal® Kit <i>Chlamydiaceae</i> order no. DVEB03111 or DVEB03151	4
5.2. BactoReal® Kit <i>Chlamydiaceae</i> order no. DVEB03113 or DVEB03153.....	4
6. Additionally required materials and devices.....	4
7. Preparation of real-time PCR.....	5
7.1. Pipetting scheme	5
7.2. Programming of the temperature profile.....	5
8. Interpretation of PCR-data	6
9. Troubleshooting.....	8
10. Specifications	8
10.1. Analytical sensitivity.....	8
10.2. Analytical specificity.....	8
11. Annex – symbols.....	8

1. Product description

BactoReal® Kit *Chlamydiaceae* is a real-time PCR assay for detection of DNA of *Chlamydia* and *Chlamydophila* species. This test was developed and validated for the ABI PRISM® 7500 (Fast) instrument (Life Technologies), LightCycler® 480 (Roche) and Mx3005P® (Agilent), but is also suitable for other real-time PCR instruments. This test allows the rapid and sensitive detection of DNA of *Chlamydia* and *Chlamydophila* species from DNA samples purified from different sample material (e.g. with the QIAamp DNA Mini Kit).

BactoReal® Kit *Chlamydiaceae* detects the 23S rRNA gene of *C. muridarum*, *C. suis*, *C. trachomatis*, *C. abortus*, *C. caviae*, *C. felis*, *C. pecorum*, *C. pneumoniae* and *C. psittaci*. A probe-specific amplification-curve at 530 nm (FAM channel) indicates the amplification of *Chlamydiaceae* specific DNA.

An internal positive control system for detection in VIC/HEX channel, (554 nm, order no. DVEB03111 or DVEB03151) or Cy5 channel (667 nm; order no. DVEB03113 or DVEB03153) excludes false-negative interpretation of results due to inhibition of real-time PCR (see 8. Interpretation of PCR-data).

For subtyping of *Chlamydiaceae* the PCR-product can be sequenced and BLAST-analysed. Please contact ingenetix to request for sequencing-primer and further information.

When using PCR-platforms not validated by ingenetix, an evaluation of the multiplex-PCR is recommended. Please be aware that some PCR-platforms have to be calibrated with the corresponding dye before performing multiplex-PCR.

BactoReal®, MycoReal, ParoReal and ViroReal® Kits are optimized to run under the same thermal cycling conditions. RNA and DNA material can be analysed in one run.

2. Pathogen information

The *Chlamydiaceae* family currently includes two genera and one candidate genus: genus *Chlamydia* (including the species *Chlamydia muridarum*, *Chlamydia suis*, *Chlamydia trachomatis*), genus *Chlamydophila* (including the species *Chlamydophila abortus*, *Chlamydophila caviae*, *Chlamydophila felis*, *Chlamydia pecorum*, *Chlamydophila pneumoniae*, *Chlamydophila psittaci*), and candidatus *Clavochlamydia*. All *Chlamydiaceae* are obligate intracellular Gram-negative bacteria that cause diseases in humans and animals worldwide.

References:

Everett, K. D. E., Bush, R. M. & Andersen, A. A. 1999. Emended description of the order *Chlamydiales*, proposal of *Parachlamydiaceae* fam. nov. and *Simkaniaceae* fam. nov., each containing one monotypic genus, revised taxonomy of the family *Chlamydiaceae*, including a new genus and five new species, and standards for the identification of organisms. *International Journal of Systematic and Evolutionary Bacteriology* 49, 415 - 440.

3. Principle of real-time PCR

A specific DNA sequence of the pathogen genome is amplified and the generated PCR-product is detected by an oligonucleotide-probe labelled with a fluorescent dye. This technology allows for a sequence-specific detection of PCR amplicates.

4. General Precautions

The user should always pay attention to the following:

- Always include a negative control per PCR-run (water instead of sample).
- Optional: for valid interpretation of results, a negative control should be included during DNA-extraction (for example extraction of water instead of sample material), in order to exclude false-positive results due to contamination with *Chlamydiaceae* DNA during extraction.
- Be careful when handling the positive control.
- Store and extract positive material (specimens, controls and amplicons) separately from all other reagents and add it to the reaction mix in a spatially separated workspace.
- Periodically decontaminate benches and devices.
- Use sterile pipette tips with filters.
- Thaw all components thoroughly at room temperature before starting an assay. When thawed, mix the components and centrifuge briefly.
- For MSDS, see www.ingenetix.com.

5. Contents of the Kit

5.1. BactoReal® Kit *Chlamydiaceae* order no. DVEB03111 or DVEB03151

Labelling	Content	Amount		Storage
		DVEB03111	DVEB03151	
<i>Chlamydiaceae</i> Assay Mix (green cap)	Primer and probe (FAM) for detection of <i>Chlamydiaceae</i>	2 x 50 µl	1 x 50 µl	-20°C
CR-1 Assay Mix (yellow cap)	Primer, probe (VIC/HEX) and target for detection of IPC	2 x 50 µl	1 x 50 µl	-20°C
<i>Chlamydiaceae</i> Positive Control (red cap)*	Control-DNA (approx. 10,000 target copies/µl)	1 x 25 µl	1 x 25 µl	-20°C
DNA Reaction Mix (white cap)#	Reaction Mix	2 x 500 µl	1 x 500 µl	-20°C until first use, then at +4°C
Water (blue cap)	Water	1 x 1000 µl	1 x 1000 µl	-20°C to +4°C

#DNA Reaction Mix contains uracil-N glycosylase (UNG)

5.2. BactoReal® Kit *Chlamydiaceae* order no. DVEB03113 or DVEB03153

Labelling	Content	Amount		Storage
		DVEB03113	DVEB03153	
<i>Chlamydiaceae</i> Assay Mix (green cap)	Primer and probe (FAM) for detection of <i>Chlamydiaceae</i>	2 x 50 µl	1 x 50 µl	-20°C
CR-3 Assay Mix (yellow cap)	Primer, probe (Cy5) and target for detection of IPC	2 x 50 µl	1 x 50 µl	-20°C
<i>Chlamydiaceae</i> Positive Control (red cap)*	Control-DNA (approx. 10,000 target copies/µl)	1 x 25 µl	1 x 25 µl	-20°C
DNA Reaction Mix (white cap)#	Reaction Mix	2 x 500 µl	1 x 500 µl	-20°C until first use, then at +4°C
Water (blue cap)	Water	1 x 1000 µl	1 x 1000 µl	-20°C to +4°C

#DNA Reaction Mix contains uracil-N glycosylase (UNG)

The components of BactoReal® Kit *Chlamydiaceae* are stable until the expiry date stated on the label. Repeated thawing and freezing should be avoided. Please protect kit components from light.

6. Additionally required materials and devices

- Reagents and devices for DNA-extraction
- PCR-grade water
- Disposable powder-free gloves
- Pipettes (adjustable)
- Sterile pipette tips with filters
- Vortex mixer
- Desktop centrifuge with rotor for 2 ml reaction tubes
- Real-time PCR instrument which is able to detect and differentiate fluorescence in FAM and VIC/HEX or Cy5 channel
- Appropriate 96 well reaction plates or reaction tubes with corresponding (optical) closing material

7. Preparation of real-time PCR

Please make sure that at least one negative control (water, blue cap), as well as one positive control (red cap) and one extraction negative control (optional, recommended) are included per PCR run.

Ingenetix highly recommends performing PCR analyses in duplicates, which increases the probability of detection of the pathogen and facilitates interpretation of results.

7.1. Pipetting scheme

		Per sample
Preparation of Master Mix (mix well)	Water*	3.0 µl
	DNA Reaction Mix (2x)	10.0 µl
	<i>Chlamydiaceae</i> Assay Mix	1.0 µl
	CR Assay Mix	1.0 µl
	Total volume Master Mix	15.0 µl
Preparation of PCR	Master Mix	15.0 µl
	Sample*	5.0 µl
	Total volume	20.0 µl

*1-8 µl of the sample can be used. When using an amount other than 5 µl of the sample, the amount of H₂O has to be changed accordingly.

Positive Control: As positive control use 1 µl of the *Chlamydiaceae* Positive Control + 4 µl H₂O.

Optional: a 1:10 dilution of the positive control can be used and defined as second standard value (approx. 1000 target copies/µl).

7.2. Programming of the temperature profile

Please find further information on programming the real-time PCR instrument in the respective operator's manual. Please be aware that some PCR-platforms have to be calibrated with the corresponding dye before performing multiplex-PCR.

Select dyes: FAM-TAMRA for detection of *Chlamydiaceae*

Cy5-NONE (CR-3 Assay Mix) or VIC-TAMRA (CR-1 Assay Mix) for detection of IPC

Select reference dye (passive reference): ROX

Sample Volume: 20 µl

Temperature Profile:

Program 1	Program 2	Program 3
Cycles: 1 Analysis: None	Cycles: 1 Analysis: None	Cycles: 45 Analysis: Quantification Acquisition at 60°
50°C 2 min*	95°C 20 sec	95°C 5 sec
		60°C 1 min

For ABI PRISM® 7500:

Ramp speed: Without "fast cycling" parameter

For LightCycler® 480 instrument:

Detection format: 2 Color Hydrolysis Probe
(dyes see above)

***Note:** If viral RNA should be also detected in the same PCR run, program 1 has to be prolonged to 15 min at 50°C. This temperature profile can be used for all BactoReal®, MycoReal®, ParoReal® and ViroReal® kits for the detection of DNA or RNA.

8. Interpretation of PCR-data

Examples for interpretation of positive reactions are shown in the amplification plots below.

For a valid interpretation, the following criteria must be fulfilled:

	Ct/Cp (FAM channel) <i>Chlamydiaceae</i> target	Ct/Cp IPC target	Interpretation
Negative control	Negative	36.0 ± 2	Valid
Positive control (undiluted, 1 µl/PCR)	28.0-31.0	36.0 ± 2	Valid
Or: positive control (1:10 diluted, 1 µl/PCR)	31.0-34.0	36.0 ± 2	Valid
Extraction negative control (optional)	Negative	36.0 ± 2	Valid
Negative sample	Negative	36.0 ± 2	Valid
Positive sample	Positive	Positive/Negative	Valid

For analysis of PCR data please proceed as follows:

For analysis of PCR results gained with BactoReal® Kit *Chlamydiaceae* please select fluorescence display options FAM channel for the *Chlamydiaceae* target and VIC/HEX channel (order no. DVEB03111, DVEB03151) or Cy5 channel (order no. DVEB03113, DVEB03153) for the internal positive control target. Samples with a positive Cp or Ct-value are considered positive. Please also check the presence of amplification-curves manually.

Once the analysis is completed, the following results are possible:

1. Signal in FAM channel:

→ DNA of *Chlamydia* or *Chlamydophila* species was amplified. The sample has to be interpreted as positive. DNA of *Chlamydia* or *Chlamydophila* species can lead to a reduced or absent fluorescence signal of the internal positive control (competition of PCR).

2. No signal in FAM channel:

→ No DNA of *Chlamydia* or *Chlamydophila* species is detectable in the sample. The sample has to be interpreted as negative.
An inhibition of PCR cannot be excluded.

2a. No signal in FAM channel but signal of the internal positive control:

→ No DNA of *Chlamydia* or *Chlamydophila* species is detectable in the sample. The sample has to be interpreted as negative.
The positive signal of the internal positive control assay excludes a putative PCR inhibition.

2b. No signals in FAM channel and no signal with internal positive control:

→ No interpretation statement can be made.
Information about possible sources of error and their solution can be found in 9. Troubleshooting.

<p style="text-align: center;">Detection of <i>Chlamydiaceae</i></p> <p style="text-align: center;">Amplification Plot</p> <p>ABI Prism® 7500: FAM channel, 530 nm 1:10 serial dilution of <i>Chlamydia</i> DNA</p>	<p style="text-align: center;">Detection of internal positive control CR-3</p> <p style="text-align: center;">Amplification Plot</p> <p>ABI Prism® 7500: Cy5 channel, 667 nm Internal positive control</p>	<p style="text-align: center;">Detection of internal positive control CR-1</p> <p style="text-align: center;">Amplification Plot</p> <p>ABI Prism® 7500: VIC channel, 554 nm Internal positive control</p>
<p style="text-align: center;">Mx3005P®: FAM channel</p> <p style="text-align: center;">Amplification Plot</p> <p>1:10 serial dilution of <i>Chlamydia</i> DNA</p>	<p style="text-align: center;">Mx3005P®: CY5 channel</p> <p style="text-align: center;">Amplification Plot</p> <p>Internal positive control</p>	<p style="text-align: center;">Mx3005P®: HEX channel</p> <p style="text-align: center;">Amplification Plot</p> <p>Internal positive control</p>
<p style="text-align: center;">LightCycler® 480: FAM channel</p> <p style="text-align: center;">Amplification Curves</p> <p>1:10 serial dilution of <i>Chlamydia</i> DNA</p>	<p style="text-align: center;">LightCycler® 480: Cy5 channel</p> <p style="text-align: center;">Amplification Curves</p> <p>Internal positive control</p>	

9. Troubleshooting

1. No *Chlamydia* or *Chlamydomphila* species specific signal with positive control:

- Incorrect programming of the temperature profile of the real-time PCR instrument.
→ Compare the temperature profile with the protocol (see 7. Preparation of real-time PCR).
- Incorrect configuration of the PCR reaction.
→ Check your work steps (see 7. Preparation of real-time PCR) and repeat the PCR, if necessary.

2. No signal with the internal positive control and no *Chlamydia* or *Chlamydomphila* species specific signal with the sample:

- The PCR reaction was inhibited. No interpretation can be made.
→ Make sure that you use a recommended method for DNA isolation and stick closely to the manufacturer's instructions.
→ If no operating mistakes during extractions can be retraced, it is recommended to repeat the PCR with lower amounts of DNA-eluate (1/5 or 1/10 of sample volume + the adequate amount of H₂O).
- Incorrect PCR conditions.
→ Check the PCR conditions and repeat the PCR, if necessary.

3. *Chlamydia* or *Chlamydomphila* species specific signal with the negative control:

- A contamination occurred during preparation of the PCR.
→ Repeat the PCR with new reagents in replicates.
→ Strictly pipette the positive controls at last.
→ Make sure that work space and instruments are decontaminated at regular intervals.

4. *Chlamydia* or *Chlamydomphila* species specific signal with the negative control of DNA-extraction:

- A contamination occurred during extraction.
→ Repeat the extraction and PCR using new reagents.
→ Make sure that work space and instruments are decontaminated at regular intervals.

10. Specifications

BactoReal[®] Kit *Chlamydiaceae* was evaluated with the ABI PRISM[®] 7500 (Fast) instrument (Life Technologies), with the LightCycler[®] 480 (Roche) and the Mx3005P[®] (Agilent). For further validation data please contact ingenetix.

10.1. Analytical sensitivity

BactoReal[®] Kit *Chlamydiaceae* detects at least 10 copies/reaction. The limit of detection (LoD₉₅ = smallest number of copies of target DNA which can be detected in 95% of cases) is 73 target copies/reaction and was determined by several replicates around the detection limit.

10.2. Analytical specificity

The specificity is ensured by the selection of highly specific primers and probes. The primers and probes were checked for possible homologies to currently published sequences by sequence comparison analyses. This also validated the detection of so far known *Chlamydia* or *Chlamydomphila* strains. Specificity was tested on *Streptococcus dysgalactiae*, *Streptococcus agalactiae*, *Listeria monocytogenes*, *Mycoplasma bovis*, *Bordetella bronchiseptica*, *Haemophilus parasuis* and *Leptospira interrogans*. No cross-reactivity was observed. This test detects *Chlamydia* and *Chlamydomphila* species.

11. Annex – symbols



Batch code



Use by



Catalogue number



Manufactured by



Contains sufficient for <n> tests



Store at