

# BactoReal<sup>®</sup> Kit *Brachyspira pilosicoli*

## Manual

For use with the

- ABI PRISM<sup>®</sup> 7500 (Fast)
- Mx3005P<sup>®</sup>
- LightCycler<sup>®</sup> 480



For veterinary use only



DVEB01211, DVEB01213



100



DVEB01251, DVEB01253



50



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**Index**

- 1. Product description ..... 3
- 2. Pathogen information ..... 3
- 3. Principle of real-time PCR..... 3
- 4. General Precautions ..... 3
- 5. Contents of the Kit..... 4
  - 5.1. BactoReal® Kit *Brachyspira pilosicoli* order no. DVEB01211 or DVEB01251 ..... 4
  - 5.2. BactoReal® Kit *Brachyspira pilosicoli* order no. DVEB01213 or DVEB01253 ..... 4
- 6. Additionally required materials and devices ..... 4
- 7. Preparation of real-time PCR ..... 5
  - 7.1. Pipetting scheme ..... 5
  - 7.2. Programming of the temperature profile ..... 5
- 8. Interpretation of PCR-data ..... 6
- 9. Troubleshooting..... 8
- 10. Specifications ..... 8
  - 10.1. Analytical sensitivity ..... 8
  - 10.2. Analytical specificity ..... 8
- 11. Annex – symbols..... 8

## 1. Product description

BactoReal® Kit *Brachyspira pilosicoli* is a real-time PCR assay for detection of *B. pilosicoli* DNA. This test was developed and validated for the ABI PRISM® 7500 (Fast) instrument (Life Technologies), LightCycler® 480 (Roche) and Mx3005P® (Agilent), but is also suitable for other real-time PCR instruments. This test allows the rapid and sensitive detection of DNA of *B. pilosicoli* from samples purified from faecal samples, or biopsies of the intestinal mucosa. *Brachyspira pilosicoli* DNA can be recovered efficiently from faecal samples using the QIAamp DNA Stool Mini Kit, and from biopsies using the QIAamp DNA Mini Kit extraction methods, for example.

BactoReal® Kit *Brachyspira pilosicoli* detects the *nox* gene of *B. pilosicoli*. A probe-specific amplification-curve at 530 nm (FAM channel) indicates the amplification of *B. pilosicoli* specific DNA.

An internal positive control system for detection in VIC/HEX channel, (554 nm, order no. DVEB01211 or DVEB01251) or Cy5 channel (667 nm; order no. DVEB01213 or DVEB01253) excludes false-negative interpretation of results due to inhibition of real-time PCR (see 8. Interpretation of PCR-data).

When using PCR-platforms not validated by ingenetix, an evaluation of the multiplex-PCR is recommended. Please be aware that some PCR-platforms have to be calibrated with the corresponding dye before performing multiplex-PCR.

BactoReal®, MycoReal, ParoReal and ViroReal® Kits are optimized to run under the same thermal cycling conditions. RNA and DNA material can be analysed in one run.

## 2. Pathogen information

*Brachyspira pilosicoli* (previously called *Serpulina pilosicoli*) is an anaerobic intestinal spirochaete. This bacterium colonizes the large intestine of various species, including humans. It is the etiologic agent of human and animal intestinal spirochetosis and is rarely implicated as a cause of bacteremia. For example, it is the causative agent of porcine colonic spirochaetosis (PCS). *Brachyspira pilosicoli* is an anaerobic bacterium but is aerotolerant due, at least in part, to high NADH oxidase activity.

### References:

Atyeo, R. F., T. B. Stanton, N. S. Jensen, D. S. Suriyaarachichi, and D. J. Hampson. 1999. Differentiation of *Serpulina* species by NADH oxidase gene (*nox*) sequence comparisons and *nox*-based polymerase chain reaction tests. *Vet. Microbiol.* 67:47–60.

## 3. Principle of real-time PCR

A specific DNA sequence of the pathogen genome is amplified and the generated PCR-product is detected by an oligonucleotide-probe labelled with a fluorescent dye. This technology allows for a sequence-specific detection of PCR amplicates.

## 4. General Precautions

The user should always pay attention to the following:

- Always include a negative control per PCR-run (water instead of sample).
- Optional: for valid interpretation of results, a negative control should be included during DNA-extraction (for example extraction of water instead of sample material), in order to exclude false-positive results due to contamination with *B. pilosicoli* DNA during extraction.
- Be careful when handling the positive control.
- Store and extract positive material (specimens, controls and amplicons) separately from all other reagents and add it to the reaction mix in a spatially separated workspace.
- Periodically decontaminate benches and devices.
- Use sterile pipette tips with filters.
- Thaw all components thoroughly at room temperature before starting an assay. When thawed, mix the components and centrifuge briefly.
- For MSDS, see [www.ingenetix.com](http://www.ingenetix.com).

## 5. Contents of the Kit

### 5.1. BactoReal® Kit *Brachyspira pilosicoli* order no. DVEB01211 or DVEB01251

Labelling	Content	Amount		Storage
		DVEB01211	DVEB01251	
<i>Brachyspira pilosicoli</i> Assay Mix (green cap)	Primer and probe (FAM) for detection of <i>B. pilosicoli</i>	2 x 50 µl	1 x 50 µl	-20°C
CR-1 Assay Mix (yellow cap)	Primer, probe (VIC/HEX) and target for detection of IPC	2 x 50 µl	1 x 50 µl	-20°C
<i>Brachyspira pilosicoli</i> Positive Control (red cap)*	Control-DNA (approx. 10,000 target copies/µl)	1 x 25 µl	1 x 25 µl	-20°C
DNA Reaction Mix (white cap)#	Reaction Mix	2 x 500 µl	1 x 500 µl	<b>-20°C until first use, then at +4°C</b>
Water (blue cap)	Water	1 x 1000 µl	1 x 1000 µl	-20°C to +4°C

#DNA Reaction Mix contains uracil-N glycosylase (UNG)

### 5.2. BactoReal® Kit *Brachyspira pilosicoli* order no. DVEB01213 or DVEB01253

Labelling	Content	Amount		Storage
		DVEB01213	DVEB01253	
<i>Brachyspira pilosicoli</i> Assay Mix (green cap)	Primer and probe (FAM) for detection of <i>B. pilosicoli</i>	2 x 50 µl	1 x 50 µl	-20°C
CR-3 Assay Mix (yellow cap)	Primer, probe (Cy5) and target for detection of IPC	2 x 50 µl	1 x 50 µl	-20°C
<i>Brachyspira pilosicoli</i> Positive Control (red cap)*	Control-DNA (approx. 10,000 target copies/µl)	1 x 25 µl	1 x 25 µl	-20°C
DNA Reaction Mix (white cap)#	Reaction Mix	2 x 500 µl	1 x 500 µl	<b>-20°C until first use, then at +4°C</b>
Water (blue cap)	Water	1 x 1000 µl	1 x 1000 µl	-20°C to +4°C

#DNA Reaction Mix contains uracil-N glycosylase (UNG)

The components of BactoReal® Kit *Brachyspira pilosicoli* are stable until the expiry date stated on the label. Repeated thawing and freezing should be avoided. Please protect kit components from light.

## 6. Additionally required materials and devices

- Reagents and devices for DNA-extraction
- PCR-grade water
- Disposable powder-free gloves
- Pipettes (adjustable)
- Sterile pipette tips with filters
- Vortex mixer
- Desktop centrifuge with rotor for 2 ml reaction tubes
- Real-time PCR instrument which is able to detect and differentiate fluorescence in FAM and VIC/HEX or Cy5 channel
- Appropriate 96 well reaction plates or reaction tubes with corresponding (optical) closing material

## 7. Preparation of real-time PCR

Please make sure that at least one negative control (water, blue cap), as well as one positive control (red cap) and one extraction negative control (optional, recommended) are included per PCR run.

Ingenetix highly recommends performing PCR analyses in duplicates, which increases the probability of detection of the pathogen and facilitates interpretation of results.

### 7.1. Pipetting scheme

		<b>Per sample</b>
<b>Preparation of Master Mix</b> (mix well)	Water*	3.0 µl
	DNA Reaction Mix (2x)	10.0 µl
	<i>Brachyspira pilosicoli</i> Assay Mix	1.0 µl
	CR Assay Mix	1.0 µl
	<b>Total volume Master Mix</b>	<b>15.0 µl</b>
<b>Preparation of PCR</b>	Master Mix	15.0 µl
	Sample*	5.0 µl
	<b>Total volume</b>	<b>20.0 µl</b>

\*1-8 µl of the sample can be used. When using an amount other than 5 µl of the sample, the amount of H<sub>2</sub>O has to be changed accordingly.

**Positive Control:** As positive control please use 1 µl of the *Brachyspira pilosicoli* Positive Control + 4 µl H<sub>2</sub>O. Optional: a 1:10 dilution of the positive control can be used and defined as second standard value (approx. 1000 target copies/µl).

### 7.2. Programming of the temperature profile

Please find further information on programming the real-time PCR instrument in the respective operator's manual. Please be aware that some PCR-platforms have to be calibrated with the corresponding dye before performing multiplex-PCR.

**Select dyes:** FAM-TAMRA for detection of *Brachyspira pilosicoli*  
Cy5-NONE (CR-3 Assay Mix) or VIC-TAMRA (CR-1 Assay Mix) for detection of IPC

**Select reference dye (passive reference):** ROX

**Sample Volume:** 20 µl

**Temperature Profile:**

<b>Program 1</b>	<b>Program 2</b>	<b>Program 3</b>
Cycles: 1 Analysis: None	Cycles: 1 Analysis: None	Cycles: 45 Analysis: Quantification Acquisition at 60°
50°C 2 min*	95°C 20 sec	95°C 5 sec
		60°C 1 min

For ABI PRISM® 7500:  
Ramp speed: Without "fast cycling" parameter

For LightCycler® 480 instrument:  
Detection format: 2 Color Hydrolysis Probe  
(dyes see above)

**\*Note:** If viral RNA should be also detected in the same PCR run, program 1 has to be prolonged to 15 min at 50°C. This temperature profile can be used for all BactoReal®, MycoReal®, ParoReal® and ViroReal® kits for the detection of DNA or RNA.

## 8. Interpretation of PCR-data

Examples for interpretation of positive reactions are shown in the amplification plots below.

**For a valid interpretation, the following criteria must be fulfilled:**

	Ct/Cp (FAM channel) <i>B. pilosicoli</i> target	Ct/Cp IPC target	Interpretation
Negative control	Negative	36.0 ± 2	Valid
Positive control (undiluted, 1 µl/PCR)	28.0-31.0	36.0 ± 2	Valid
Or: positive control (1:10 diluted, 1 µl/PCR)	31.0-34.0	36.0 ± 2	Valid
Extraction negative control (optional)	Negative	36.0 ± 2	Valid
Negative sample	Negative	36.0 ± 2	Valid
Positive sample	Positive	Positive/Negative	Valid

**For analysis of PCR data please proceed as follows:**

For analysis of PCR results gained with BactoReal® Kit *Brachyspira pilosicoli* please select fluorescence display options FAM channel for the *B. pilosicoli* target and VIC/HEX channel (order no. DVEB01211, DVEB01251) or Cy5 channel (order no. DVEB01213, DVEB01253) for the internal positive control target. Samples with a positive Cp or Ct-value are considered positive. Please also check the presence of amplification-curves manually.

**Once the analysis is completed, the following results are possible:**

### 1. Signal in FAM channel:

→ DNA of *B. pilosicoli* was amplified. The sample has to be interpreted as positive.

*B. pilosicoli* DNA can lead to a reduced or absent fluorescence signal of the internal positive control (competition of PCR).

### 2. No signal in FAM channel:

→ No *B. pilosicoli* DNA is detectable in the sample. The sample has to be interpreted as negative. An inhibition of PCR cannot be excluded.

#### 2a. No signal in FAM channel but signal of the internal positive control:

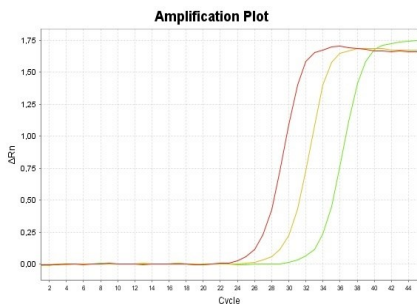
→ No *B. pilosicoli* DNA is detectable in the sample. The sample has to be interpreted as negative. The positive signal of the internal positive control assay excludes a putative PCR inhibition.

#### 2b. No signals in FAM channel and no signal with internal positive control:

→ No interpretation statement can be made.

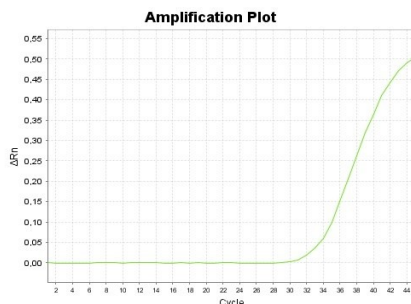
Information about possible sources of error and their solution can be found in 9. Troubleshooting.

Detection of *Brachyspira pilosicoli*



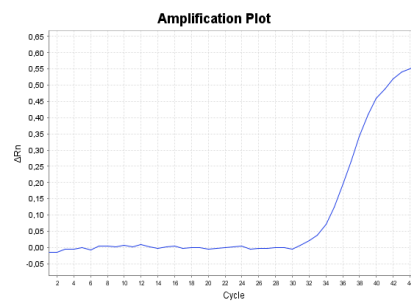
**ABI Prism® 7500:** FAM channel, 530 nm  
1:10 serial dilution of *B. pilosicoli* DNA

Detection of internal positive control CR-3

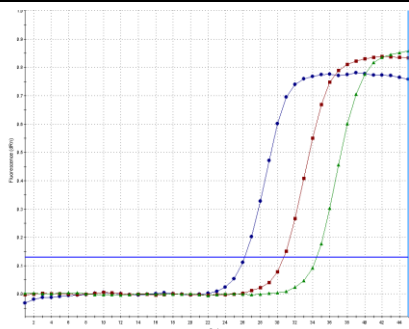


**ABI Prism® 7500:** Cy5 channel, 667 nm  
Internal positive control

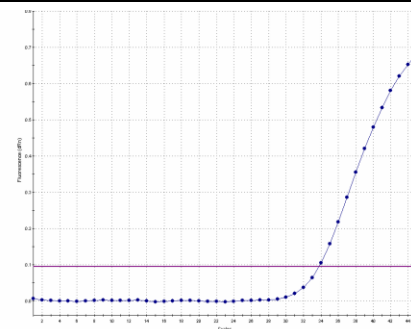
Detection of internal positive control CR-1



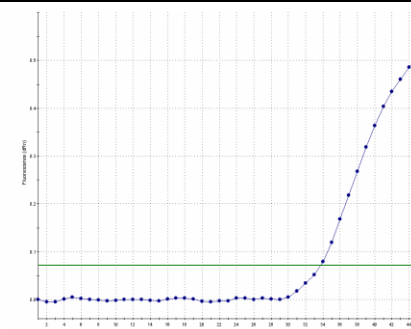
**ABI Prism® 7500:** VIC channel, 554 nm  
Internal positive control



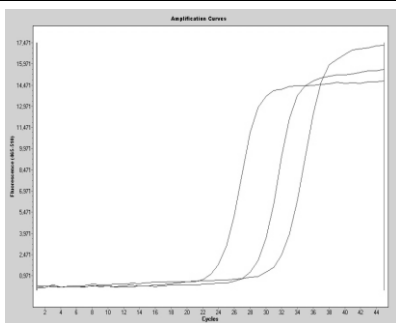
**Mx3005P®:** FAM channel  
1:10 serial dilution of *B. pilosicoli* DNA



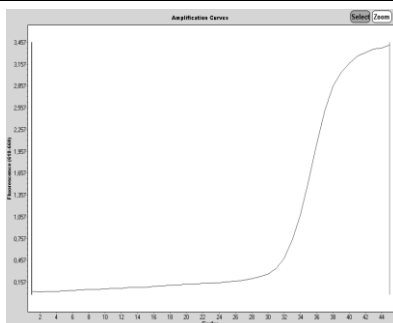
**Mx3005P®:** CY5 channel  
Internal positive control



**Mx3005P®:** HEX channel  
Internal positive control



**LightCycler® 480:** FAM channel  
1:10 serial dilution of *B. pilosicoli* DNA



**LightCycler® 480:** Cy5 channel  
Internal positive control

## 9. Troubleshooting

### 1. No *B. pilosicoli* specific signal with positive control:

- Incorrect programming of the temperature profile of the real-time PCR instrument.  
→ Compare the temperature profile with the protocol (see 7. Preparation of real-time PCR).
- Incorrect configuration of the PCR reaction.  
→ Check your work steps (see 7. Preparation of real-time PCR) and repeat the PCR, if necessary.

### 2. No signal with the internal positive control and no *B. pilosicoli* specific signal with the sample:

- The PCR reaction was inhibited. No interpretation can be made.  
→ Make sure that you use a recommended method for DNA isolation and stick closely to the manufacturer's instructions.  
→ If no operating mistakes during extractions can be retraced, it is recommended to repeat the PCR with lower amounts of DNA-eluate (1/5 or 1/10 of sample volume + the adequate amount of H<sub>2</sub>O).
- Incorrect PCR conditions.  
→ Check the PCR conditions and repeat the PCR, if necessary.

### 3. *B. pilosicoli* specific signal with the negative control:

- A contamination occurred during preparation of the PCR.  
→ Repeat the PCR with new reagents in replicates.  
→ Strictly pipette the positive controls at last.  
→ Make sure that work space and instruments are decontaminated at regular intervals.

### 4. *B. pilosicoli* specific signal with the negative control of DNA-extraction:

- A contamination occurred during extraction.  
→ Repeat the extraction and PCR using new reagents.  
→ Make sure that work space and instruments are decontaminated at regular intervals.

## 10. Specifications

BactoReal<sup>®</sup> Kit *Brachyspira pilosicoli* was evaluated with the ABI PRISM<sup>®</sup> 7500 (Fast) instrument (Life Technologies), with the LightCycler<sup>®</sup> 480 (Roche) and the Mx3005P<sup>®</sup> (Agilent). For further validation data please contact ingenetix.

### 10.1. Analytical sensitivity

The analytical sensitivity is 10 target copies/reaction. The limit of detection (LoD<sub>95</sub> = smallest number of copies of target DNA which can be detected in 95% of cases) of 25 target copies/reaction was determined by several replicates around the detection limit.

### 10.2. Analytical specificity

The specificity is ensured by the selection of highly specific primers and probes. The primers and probes were checked for possible homologies to currently published sequences by sequence comparison analyses. This also validated the detection of so far known *B. pilosicoli* strains.

BactoReal<sup>®</sup> Kit *Brachyspira pilosicoli* was tested on several different bacterial isolates (*B. hyodysenteriae*, *E. coli*, *H. parasuis*, *L. intracellularis*, *L. innocua*, *L. monocytogenes*, *P. multocida*, *S. aureus*, *S. agalactiae* and *S. pyogenes*). No cross reactions were observed. 18 field samples were tested and correctly analysed.

## 11. Annex – symbols



Batch code



Catalogue number



Contains sufficient for <n> tests



Use by



Manufactured by



Store at